



DOI: 10.5281/zenodo.46360

INVESTIGATION THE MICROBIAL DETERIORATION OF SANDSTONE FROM THE OSIRION'S SARCOPHAGUS CHAMBER AS AFFECTED BY RISING GROUND WATER LEVEL

Abdou A.O.D. El-Derby¹, Maisa M.A. Mansour² and Mohamed Z.M. Salem*³

¹Archaeology Conservation Department, Faculty of Archaeology, South Valley University, Qena, Egypt

²Conservation Department, Faculty of Archaeology, Cairo University, Giza, 12613, Egypt

³Department of Forestry and Wood Technology, Faculty of Agriculture (EL-Shatby), Alexandria University, Alexandria, Egypt

Received: 03/01/2016

Accepted: 01/03/2016

Corresponding author: zidan_forest@yahoo.com

ABSTRACT

In the present study, the microbial deterioration of sandstone from the Osirion's Sarcophagus Chamber as affected by rising ground water level was investigated by means of SEM micrograph of fungal hyphae, X-ray diffraction (XRD), and Energy-dispersive X-ray spectroscopy (EDX). The following Fungi; *Cladosporium cladosporioides*, *Aspergillus terreus*, *Curvularia lunata*, and *Acremonium falciforme* were identified on the deteriorated stone surfaces and these findings were tested by SEM investigations. The hyphae penetration of the identified microfungi caused mechanical exfoliation of building stone material as well as changing in color.

KEYWORDS: Biodegradation; Osirion's Sarcophagus; Fungi; Sandstone.

1. INTRODUCTION

Deterioration of historic buildings including chemical, physical factors, and microbial growth has been reported in many works to understanding the deterioration agents (Praderio *et al.*, 1993; Kumar and Kumar, 1999; Nuhoglu *et al.*, 2006; Suihko *et al.*, 2007). The colonization of stones by microbes depends on environmental factors such as water availability, pH, climatic exposure, and nutrient sources, and they can cause various damages on the stone surface, such as: formation of biofilm, chemical reactions with substrate, physical penetration into the substrate as well as pigments production (Pochon and Jaton, 1968; Bock and Sand, 1993; May *et al.*, 1993).

A considerable number of investigations have been examined the essential role of biological agents in the deterioration of stone (Saiz-Jimenez 1997; Tomaselli *et al.*, 2000; Warscheid and Braams, 2000; Zanardini *et al.*, 2000; Herrera *et al.*, 2004). For example, whole surface of monuments stone under air pollution and continental-cold climatic conditions in Erzurum, Turkey, was covered with a biofilm caused by the microorganisms (Nuhoglu *et al.*, 2006). Discoloration and degradation of different types of stone in cultural heritage objects could be caused by the fungal ability in production of pigments and organic acids (Gupta and Sharma, 2012). Additionally, the major biodeteriogens of the historic monuments and artworks are summarized in the review article of Dakal and Cameotra (2012).

Fungal Strains of the genera of *Penicillium* and *Fusarium* which excreted oxalic, fumaric and succinic acids with corrosive effects on the stony materials were observed in the decayed sandstone from the church of Carrascosa del Campo (Spain) (Gómez-Alarcón *et al.* 1995a). From both granite and sandstone substrata, the following fungi were found; *Alternaria* sp. 1,2, *Cladosporium cladosporioides*, *C. sphaerospermum*, *Epicoccum purpurascens*, *Fusarium* sp., *Mycelia sterilia* (melanised), and *M. sterilia* (non-melanised) (Ljaljević-Grbić and Vukojević, 2009). Also, the formation of extracellular melanin-polysaccharide stable complexes was the main role of *Ulocladium atrum* in biodeterioration of stone (Gutiérrez *et al.* 1995). It was observed that *Aspergillus Scalrotium* was found as dominant in the deteriorated stones of Mahadev temple, Bastar, Chhatisgarh (Gupta and Sharma, 2012). The most representative genera found in the deteriorated stonework of the fountain of Bibatauin in Granada, Spain, were *Cosmarium*, *Phormidium* and *Symploca*, and the main mineral was calcite (Zurita *et al.* (2005).

SEM-EDX, FT-IR and XRD have been used by almost all investigators in analytical techniques for

biodeterioration studies and for all types of substrate of construction materials (Gómez-Alarcón *et al.* 1995a). Silicon, aluminum, calcium, potassium, titanium, magnesium, zinc, sulfur, iron, sodium, and niobium were found in the stones of the historical buildings with varying amounts through the SEM-EDS analysis (Nuhoglu *et al.*, 2006).

The aim of this work was to evaluate the potential damage caused by fungal community on biodeteriorated stone samples from the Osirion's Sarcophagus Chamber as affected by rising ground water level by means of SEM micrograph of fungal hyphae, X-ray diffraction (XRD), and Energy-dispersive X-ray spectroscopy (EDX).

2. MATERIALS AND METHODS

2.1 Site Description

The sarcophagus chamber in the shape a huge sarcophagus or the sarcophagus of Osiris-Seti (Figure 1) is a part of the Osireion (named after and for the god Osiris), the Osireion is unfinished unique subterranean water-based nature (to represent the primeval mound of creation surrounded with the primeval waters or the waters of Nun) structure, excavated in Thebes/Eсна shale formations, directly to the west and on the same axis of Seti I temple at Abydos, the sarcophagus chamber is a rectangular room (27.20 × 4.75 × 7.55 m high), located to the east of the great central hall, it corresponds and parallels to a transverse room or hall (Entrance Hall), it was built from local limestone (from Thebes and transitional Drunka formations) with Nubian sandstone saddle roof of Nubian sandstone retains on its surface a fascinating religious, funerary and astronomical inscriptions and scenes, especially on the north side where the sky goddess Nut stretched out and bended over the world while the sun is going in the heart of the sky (El-Derby, 2005).

The sarcophagus chamber has been suffering from rising ground water level which reaches about 1.15 m. above its ground level, and the lack of the ventilation openings has led to a high relative humidity to more than 90% inside it (in its microclimate), resulting in presence, growth and activity of microorganisms on the surfaces and inside the walls of the chamber. The primary source of ground water was formerly the Nile River, where a pipe was designed to provide with water from a Nile branch, which was up to the area.

The ground water source at Osirion is the high ground water level, which rose as a result of several reasons: (a) the irrigation methods in the region since has changed 1942, (b) the construction of the High Dam and the leakage of water from Lake Nasser to (c) Nubian reservoir and from there to the other sub-

tanks, including the reservoirs located below the Osirion area, (d) the topography of the region, where the flow of groundwater from west to east changing the course from the south to the north east - after colliding Eocene limestone tongue which mediates the two bays caught west both of Nag Hammadi and Abydos and in the west of Abydos about 1 km which acts as a hydrological water barrier groundwater, inasmuch as the limestone plateau is in port, rebounding eastward and passes down Osirion leading to high water table constantly, and (e) the water flow from west to east often be the free water (a volatile water) attributed resulting from sewage, agricultural drainage water, drinking water and leaking from the soles of the irrigation and drainage channels, which linked directly or indirectly with the Nile River and affected by it and they are one of the important sources of raising the ground water level. The Influential primary reservoir of the region is the free reservoir; it is wobbling and growing steadily tank.

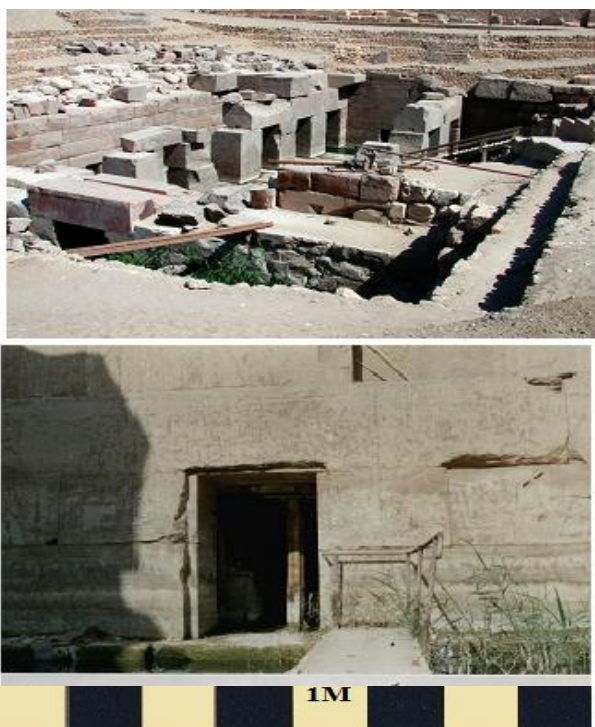


Figure 1. General view of Osirion's Sarcophagus Chamber.

2.2 Sampling Site

Biological samples of Osirion sandstone (Figure 2) were collected from different spots of the temple surface that showing decay signs such as color change and typical microbial growths. Samples were taken for mycological analyses by swabbing surfaces with sterile cotton swabs. The samples were then stored at 4°C.



Figure 2. Osirion sandstone sampling areas (A), (B), (C) and (D).

2.3 X-ray Diffraction Analysis of Stone

The chemical composition of Osirion stones was analyzed by X-ray diffraction (XRD) using Philips-Eindhoven Compact X-ray Diffractometer PW 1840. The identification of chemical compositions was carried out automatically using mineralogical data base.

2.4 Microscopic examination

Microscopic examination was performed using Leica Zoom 2000 microscope equipped with an Olympus e-410 digital camera. Thin sections, cut across the thickness of the stone, were used to determine the different typologies of sandstone.

2.5 Isolated microorganisms from the stone Temple

Microorganisms' isolation was performed directly in the laboratory after swabbing. The fungi were isolated by rubbing the swabs gently on different culture media. The media comprised 1-M40Y; 400g sucrose, 20g Molt extract, 5g Yeast extract, 20g Agar; 2- potato-dextrose agar (PDA); 200g Potato extract, 20g Agar and 20 g dextrose. Inoculated petri dishes were incubated at $25\pm 2^{\circ}\text{C}$ for fungi. Resulted cultures were purified using the hyphal tip and/or a single spore technique as described by Hildebrand (1938) and Smith and Onions (1994). Macroscopic and microscopic characteristics of the obtained isolates, as well as color, size and morphology of the vegetative and reproductive structures were examined. Fungal isolates that speculated were identified using the taxonomic keys of Ellis and Ellis (1976), and Samson *et al.*, (2010).

2.6 Colonization Test

The stone samples were cut into cubes (10 x 20 x 40 mm) using a low-speed diamond saw. The cubes' surfaces were left unpolished in order to increase surface roughness and thus facilitate microbial colonization (Guillitte, 1995). Cubes were sterilized using UV light exposure for 48 hours, after autoclaving at 121°C for 6 hours and drying in an oven at 115°C for 24 hours (Miller *et al.* 2009). For the preparation of spore suspensions, 10 ml of sterilized distilled water was added to culture plates containing PDA (7-days old), and then spores were freed by the aid of a camel brush. Afterwards spore suspensions were individually through muslin and standardized to contain 1.2×10^6 spores per ml using a haemocytometer slide. Stone cubes were deliberately inoculated by fungal and bacterial species to study the stone blocks before and after infection. The colonization was evaluated after three months' period. Degradation occurred were determined on the surface and inside the pores.

SEM model-a FEI Quanta 200 SEM FEG was used to study colonization of the stone surface by microorganisms. ESEM attached with electron dispersive X-ray spectroscopy (EDX) was used for the elemental analysis characterization of both the inoculated and non-inoculated stone sample (Danilatos and Robinson, 1979). EDX analysis was used to study changes in surface morphology and particle size of the deteriorated samples.

3. RESULTS AND DISCUSSION

3.1 X-ray diffraction analysis of stone

Result of X-ray diffraction analysis is shown in figure 3. The interpretation of the diffraction patterns indicated that the stone temple is mainly composed of silica. Also, the carbonate was found in form of Cement (Calcareous Sandstone or as biogenic carbonate precipitated as a result of dissolution of Carbonate of the adjacent walls).

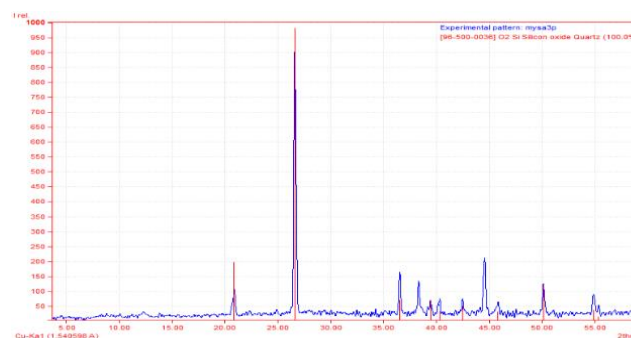


Figure 3. X-ray diffraction pattern of the sandstone

3.2 Microscopic Examination of Sandstone

Photomicrograph of sandstone thin section of quartzite (silicified sandstone) presented in figure 4, revealed the characteristic silica grains of sandstone. Due to a basalt-intrusion the interstitial space of the Oligocene sand grains is filled up with small quartz crystals precipitated from induced hydrothermal fluids.

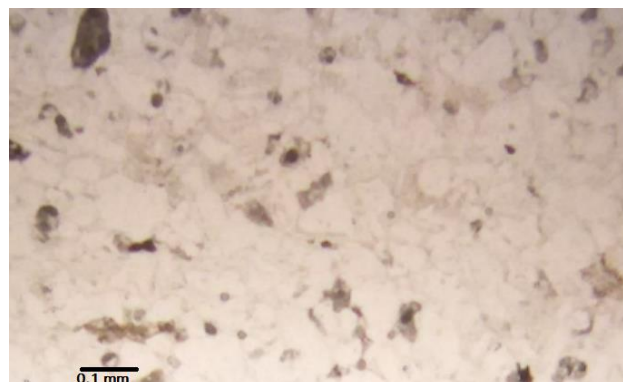


Figure 4. Photomicrograph of sandstone thin section from the site of sarcophagus of Osirion.

According to SEM examination, the original sample in Fig. 5 (SEM micrograph X 300) of Nubian Sandstone of Sarcophagus Chamber of Osirion showing existence of kaolinite either as a binding material or as a result of alteration of K-feldspar to clay minerals (kaolinite) (Kaolintization), micro-exfoliation of K-feldspar, Sodium Chloride (halite) salt, acicular radical crystals of gypsum salts. It has been apparent that fungi comprise a significant component of micro biota in a wide range of rocks including sandstone, granite, limestone, marble and gypsum (Warscheid and Braams, 2000; Burford et al., 2003a).

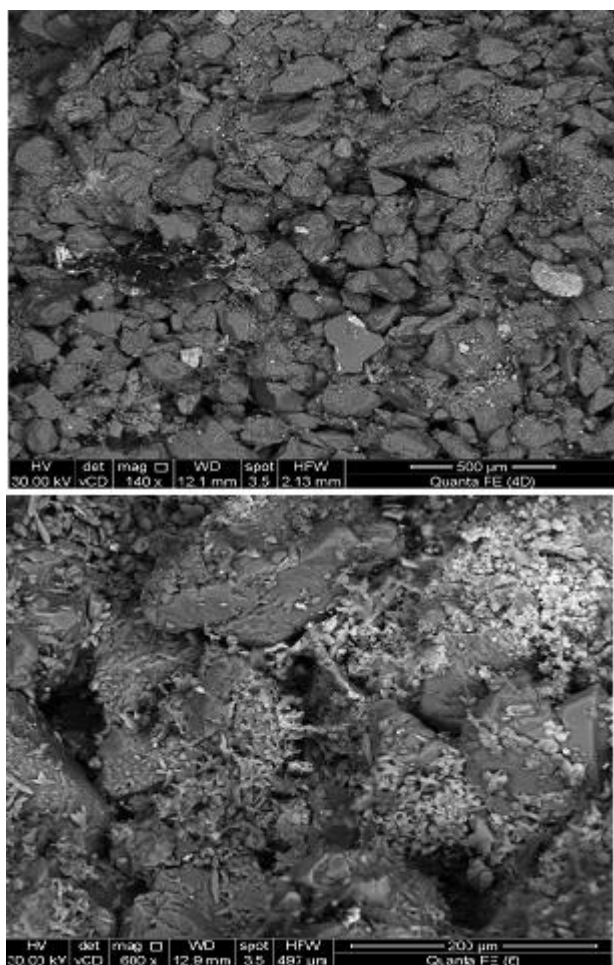


Figure 5. SEM micrograph X 1200 of Nubian Sandstone of Sarcophagus Chamber of Osirion.

Electron dispersive X-ray (Fig. 6) for check point at the same sample of weathered Nubian Sandstone surface presented high Si, O and Ca and other elemental components (Table 1) such as Cl, Fe, Al, Na, C, N, Mg and S indicating to existence salts of halite and gypsum, alteration to clay minerals and the voids between granules as a result of the loss of the binding materials (granular disintegration). Some of these elements could be used as an energy resource for the microorganisms (Nuhoglu et al., 2006). Fungi concentrated in stone crusts are able to penetrate

into the rock material by hyphal growth and biocorrosive activity, due to excretion of organic acids or by oxidation of mineral-forming cations, preferably Fe and Mg (Warscheid and Braams, 2000; Ljaljević-Grbić and Vukojević, 2009). It was also reported that the large pore sandstones promote microbial contamination only temporarily, whereas, small pore stones offer more suitable conditions for bioreceptivity (Warscheid and Braams, 2000).

In the present study, the C reached 16.81 %wt and the presence of significant amounts of carbonate compounds (e.g. > 3% w/v CaCO₃) in calcareous sandstones results in the buffering of biogenic metabolic products, also the organic adhesives, which are present in ancient brick and mortar, increase the susceptibility of the mineral substrate to microbial attack (Warscheid and Braams, 2000). It was reported that fungi can transform and weather the mineral surface of the stone by precipitating calcium carbonate into calcium oxalate by the actions of secreted oxalic acid using the process of secondary mineral formation (Burford et al., 2003b; Adeyemi AO, Gadd, 2005).

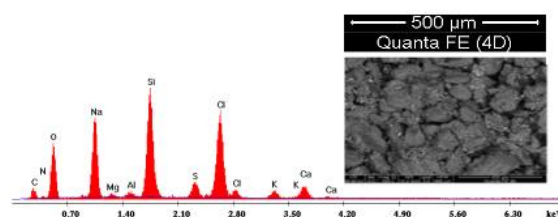


Figure 6. Electron dispersive X-ray for Elemental analysis of weathered Nubian Sandstone surface.

Table 1. Elemental composition of deteriorated sandstone

Element (Wt %)										
C	N	O	Na	Mg	Al	Si	S	Cl	K	Ca
16.81	3.62	30.17	16.95	0.68	0.80	14.33	2.18	11.42	1.16	1.89

3.3 Isolation of microorganisms from the stone temple

Isolation trials of microorganisms associated with the sandstone temple resulted in different fungal including *Cladosporium cladosporioides*, *Aspergillus terreus*, *Curvularia lunata* and *Acremonium falciforme*.

The colonization of microorganism on sandstone cubes after three month showed a growth in the cubes' surfaces. It is possible using Scanning microscopes, to identify the microorganism species through the distinctive size and shape of their spores and hyphae for fungi. SEM micrograph (Fig. 7) of Nubian Sandstone of Sarcophagus Chamber of Osirion showed gypsum crusts, fractures and microfrac-

tures in quartz grains and voids between granules as a result of the loss of the binding materials (granular disintegration).

Cladosporium cladosporioides (Fig. 8) showed small sized simple or branched loose chained spores with lemon-shape and the conidiophore forming many aerial hyphae. Gypsum crusts were recently-formed by *C. cladosporioides*, where the calcite has been converted to gypsum (from insoluble stage to soluble stage in water).

The presence of fungi on the sandstone surface of the Osirion's Sarcophagus Chamber monuments is often associated with the process of biodeterioration. Their growth on stone surface could be altered it by the excreting inorganic and organic acids as a result of their own metabolism which generates organic acids (oxalic acid and citric acid) that have chelating properties by weaken the metal-oxygen bond, and increases the solubility of some metals and forms complexes with the mineral cations present on the surface matrix (Gómez-Alarcón et al., 1995a,b; Banfield et al., 1999; Harley and Gilkes, 2000). Also, the intrusion of fungal hyphae along the crystal plane by some fungi is known to destabilize the stone texture resulting in its mechanical deterioration (Gadd, 2005).

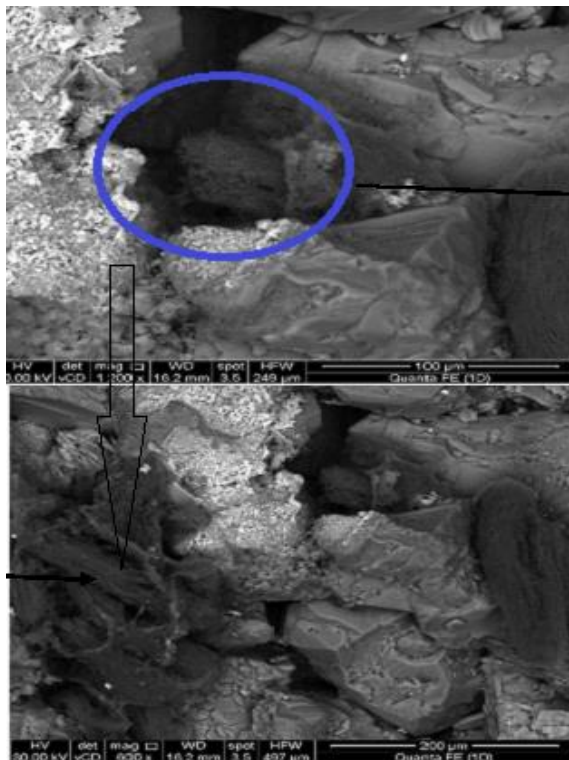


Figure 7. SEM micrograph of Nubian Sandstone of Sarcophagus Chamber of Osirion.

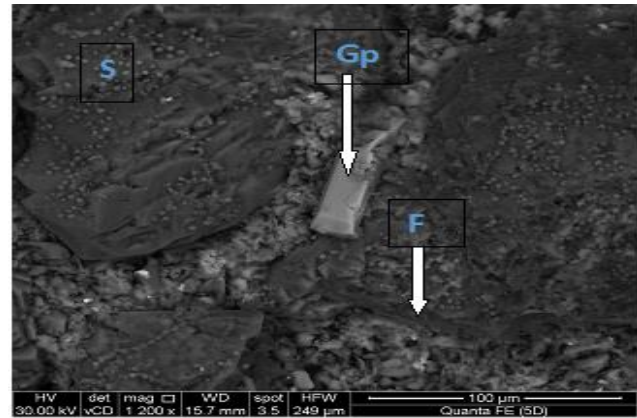


Figure 8. SEM micrograph of fungal hyphae attached to growth into sandstone after three months by *Cladosporium cladosporioides* (F), gypsum (Gp), and spore for *C. cladosporioides* (S)

SEM micrograph (Fig. 9) shows the biogenic alteration and formation of vugs filled with crystalline salts and the highly altered kaolinitic Nubian Sandstone and development of interstitial salt encrustation, vesicular, brittle sandstone and *Aspergillus terreus*.

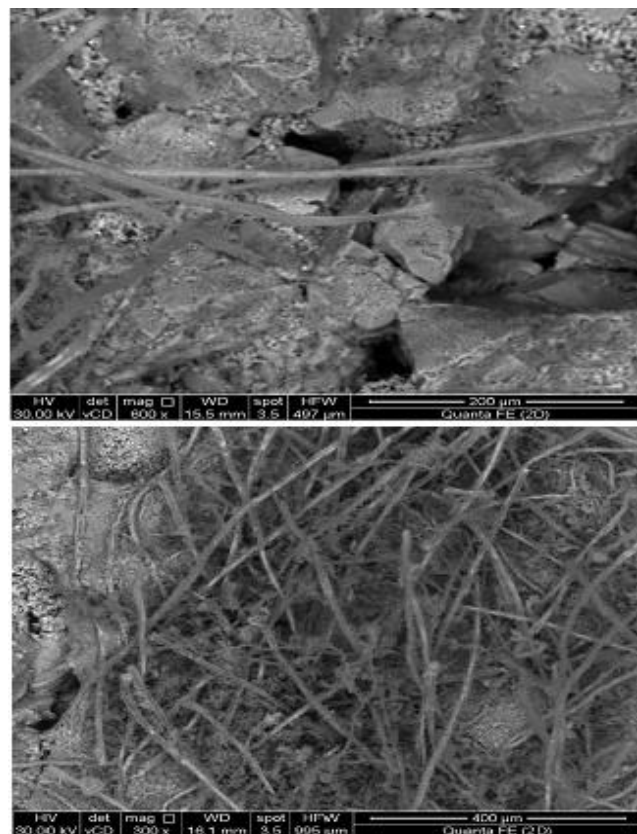


Figure 9. SEM micrograph of fungal hyphae attached to growth into sandstone after three months by *Aspergillus terreus*.

SEM micrograph (figure 10) shows the biogenic mamilated (nipples) texture of salts, brittle sandstone and *Curvularia lunata* matured colonies ap-

peared dark brown to black. Conidiophores arising from surface or aerial hyphae, septate, straight or flexuous, with apices straight, curved or conidia that are relatively short 3-septate, curved and smooth branching patterns of aerial hyphae intermingled with spore-forming structures aerial hyphae, conidiophores and spores.

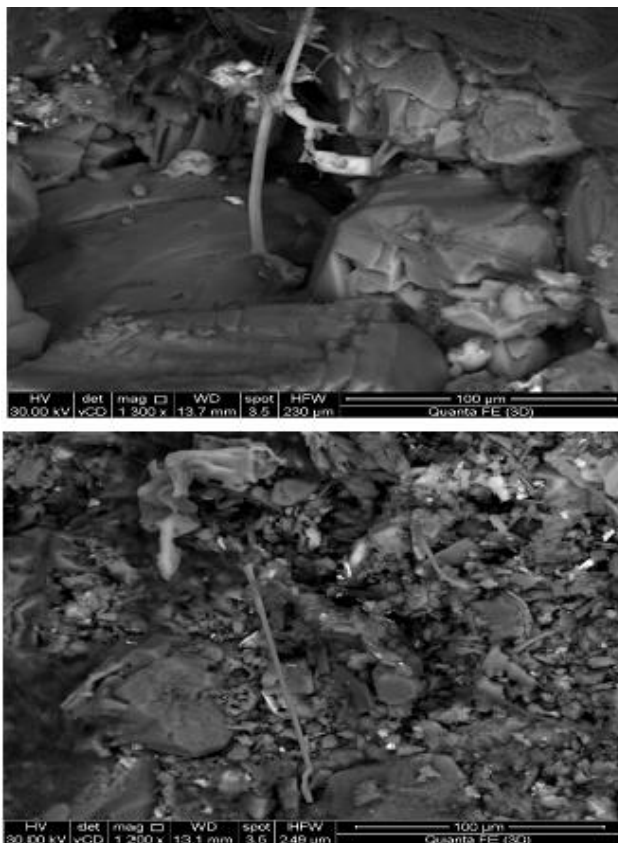


Figure 10. SEM micrograph of fungal hyphae attached to growth into sandstone after three months by *Curvularia lunata*.

SEM micrograph (Fig.11) shows the kaolinite structure tiny microbial accumulation Corroded Kaolin minerals as six sided crystals in right upper corner *Acremonium falciforme*. *A. falciforme* showed thin hyaline septate hyphae with phialides unbranched, solitary, formed on the hyphae, with the fusiform conidia.

Soil fungi, as well as air and plant-derived ora, are the first inhabitants on the dry inorganic or mineral surface in the in stone monuments which produce fast-spreading hyphal structures and can regenerate their presence on the stone by spore production (Gorbushina and Vlasov, 1998; Gorbushina and Krumbein, 2000). *C. cladosporioides* was previously found in the deteriorated sandstone surface biofilm of the Monument of the "Unknown Hero" (Avala Mountain near Belgrade) and "Brankov most" (Sava river, Belgrade) (Ljaljević-Grbić and Vukojević, 2009).

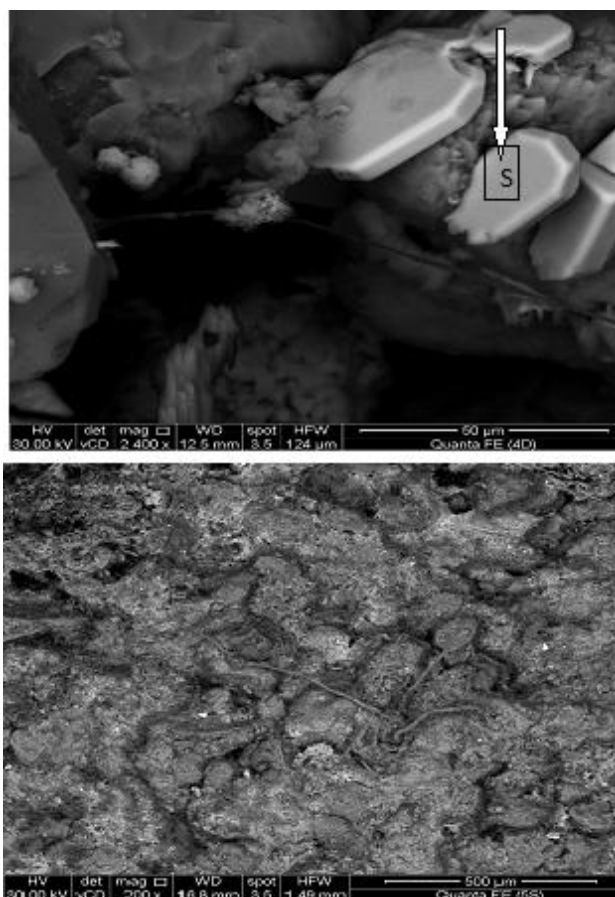


Figure 11. SEM micrograph of fungal hyphae attached to growth into sandstone after three months by *Acremonium falciforme*, Salt (S).

It was observed previously, the size of vegetative and generative organs (forms) of the microorganisms observed by the SEM images on stone surfaces are smaller than the cultural medium, soil and plant surfaces because the stone is a poorer enrichment media than the cultural medium for the microorganisms (Dornieden et al., 2000; van der Gast et al., 2001).

The elimination of sources appropriate for the growth of fungus environmental problem, control measures and reducing the groundwater tables of ground water in the Osirion and its surroundings, as follows:

1- Eradication, of agricultural, housing, urban abuses, encroachment and man-made destruction.

Encouragement and motivation Archaeological and cultural Awareness, whereas Osirion is an unique construction all over the ancient Egyptian civilization, this cultural value has not been corresponded and retaliated with care and conservation starting from arresting and staying the causes of failure, then the abstract and moral of the cultural value of Osirion will impel and inspire to removing neglecting before removing causes and symptoms of deterioration and inclusion in management site.

2- Lowering the high ground water tables with the following procedures:

(i) The regional insulation of presuppose context of the site (see: (in Arabic; El-Derby, 2005).

(ii) The local insulation of the structure and their close context.

(iii) Converting the unlined and uncovered channels and drains -to the covered subsurface drain system.

(iv) Converting the soaking perennial irrigation system to spray and dropping system.

3- Prevention procedures, such as continuous cleaning, compensation, repairing of cracks, joints, crevices and cavities.

4- Controlling the relative humidity (more than 90%) in the Chamber- which lacks windows and ventilation - with ventilation techniques and indoor air suction, and with using dehumidifier materials.

5- Installation an artificial lighting inside the Chamber, which should be appropriate in terms of kind of radiation, and in terms of aesthetics, so as to oust the bat and limit the appropriate environment for the growth of fungus.

6- Fungi control measures: it is suggested the chemical methods (herbicides) such as Glyphosate (commercial nom is Round Up), it is derived from Glycine acid and Fuluazifop-p-buty commercial nom is Fusilade) (El-Derby, 2005, in Arabic).

7- Consolidating the walls and ceiling with consolidate and water repellent, then fixing the surface scales plaster layers, which were expelled by hyphae.

Also conservation treatment was applied using UV light on sand-stone, where the UV light acts as sterilization against the growth of microorganisms.

4. CONCLUSIONS

Our results indicate that the microbial deterioration of sandstone from the Osirion's Sarcophagus Chamber as affected by rising ground water level was observed by the following fungi; *Cladosporium cladosporioides*, *Aspergillus terreus*, *Curvularia lunata*, and *Acremonium falciforme*. The results included enrichment by organic substances, formation of crusts, and chemical weathering, promote the colonization of natural sandstone by fungi and thus enhances biodegradation processes.

REFERENCES

- Adeyemi, A.O, and Gadd, G.M. (2005) Fungal degradation of calcium-, lead- and silicon-bearing minerals. *Biometals*, Vol.18, 269-281.
- Banfield JF, Barker WW, Welch SA, Taunton A (1999) Biological impact on dissolution: application of the lichen model to understanding mineral weathering in the rhizosphere. *Proc Natl Acad Sci USA*, Vol. 96, 3404-3411.
- Bock, E., Sand, W. (1993): *The microbiology of masonry biodeterioration*, J. Appl. Bacteriol. 74: 503-514.
- Burford E.P., et.al, (2003a) Fungal involvement in bioweathering and biotransformation of rocks and minerals. *Mineralogical Magazine*, Vol.67, 1127-1155.
- Burford EP, Kierans M, and Gadd GM (2003b) Geomycology: fungal growth in mineral substrata. *Mycologist*, Vol.17, 98-107.
- Dakal TC, and Cameotra SS (2012) Microbially induced deterioration of architectural heritages: routes and mechanisms involved. *Environmental Sciences Europe*, Vol.24, No.36, 1-13.
- Danilatos, G.D., and Robinson, V.N.E., (1979) Principles of scanning electron microscopy at high specimen pressures. *Scanning*, Vol.2, 72-82.
- Dornieden Th, Gorbushina A.A., and Krumbein W.E. (2000) Biodecay of cultural heritage as a space/time-related ecological situation—an evaluation of series of studies. *Int Biodeterior Biodegrad* Vol.6, 261-70.
- El-Derby, A.A.O.D. (2005) The Architectural Conservation and Maintenance of Some Ancient Egyptian Temples in Upper Egypt, An Analytical Study of Deterioration Factors & Symptoms and The Strategy for Treatment, With Application on Selected Case Study (in Arabic), A thesis for the degree of Doctor of Philosophy, submitted to Archaeology Conservation department , Faculty of Archaeology, Cairo University, 2005, pp.212-224, 285-286, figs. 26, 225.
- Ellis, M.B., and Ellis, P.J. (1997) *Microfungi on Land Plants, An Identification Handbook*, The Richmond Publishing Co., Ltd.
- Gadd GM. (2004) Mycotransformation of organic and inorganic substrates. *Mycologist*, Vol.18, 60-70.
- Gorbushina, A.A., and Vlasov, D.Y. (1998) Biodiversity of rock dwelling poikilotroph fungal communities with decreasing nutrient content of the habitat. Sixth International Mycological Congress, Jerusalem, Abstracts, p. 140.

- Gorbushina, A.A., and Krumbein, W.E., (2000) Rock dwelling fungal communities: diversity of life styles and colony structure. In: Seckbach, J. (Ed.), *Journey to Diverse Microbial Worlds. Adaptation to Exotic Environments*. Kluwer, Dordrecht, pp. 317-334.
- Gómez-Alarcón, G., Muñoz, M., Arino, X., and Ortega-Calvo, J.J. (1995a) Microbial communities in weathered sandstones: the case of Carrascosa del Campo church, Spain. *The Science of the Total Environment*, vol.167, 249-254.
- Gómez-Alarcón G, Muñoz ML, Flores M (1995b) Excretion of organic acids by fungal strains isolated from decayed sandstone. *Int Biodegrad.*, Vol.34, 169-180.
- Gutiérrez, A., Martínez, M.J., Almendros, G.C., González-Vila, and Martínez, A.T. (1995) Hyphal-sheath polysaccharides in fungal deterioration. *Sci. Total Environ.*, Vol.167, No.(1-3), 315-328.
- Guillitte, O. (1995) Bioreceptivity: A new concept for building ecology studies. *Sci. Total Environ.*, Vol.167, 215-220.
- Gupta, S.P, and Sharma, K. (2012) The role of fungi in biodeterioration of sandstone with reference to Mahadev temple, Bastar, Chhatisgarh. *Recent Research in Science and Technology* Vol.4, No.(3), 18-21.
- Harley, A.D, and Gilkes, R.J (2000) Factors influencing the release of plant nutrient elements from silicate rock powders: a geochemical overview. *Nutr Cycl Agroecosyst* Vol 56, 11-36.
- Herrera LH, Arroyave C, Guiamet P, de Saravia SG, and Videla H. (2004) Biodeterioration of peridotite and other constructional materials in a building of the Colombian Cultural Heritage. *Int Biodeterior Biodegrad.*, Vol.53, 135-41.
- Hildebrand, E.M. (1938) Techniques for the isolation of single microorganisms. *Botanical Review*, Vol.4, 627-664.
- Kumar R, and Kumar AV. (1999) In: Agnew Neville, editor. Biodeterioration of stone in tropical environments. USA7 J. Paul Getty Trust; 1999(0-89236-550-1). p. 1-2.
- Ljaljević-Grbić M, Vukojević J B. (2009) Role of fungi in biodeterioration process of stone in historic buildings. *Proc. Nat. Sci, Matica Srpska Novi Sad* Vol.116, 245-251.
- May, E., Lewis, F. J., Pereira, S., Tayler, S., Seaward, M.R.D. and Al Isopp, D. (1993) Microbial deterioration of building stone – a review, *Biodeterioration Abstracts* Vol.7, 109-123.
- Nuhoglu Y., Oguz E., Uslu H., Ozbek A., Ipekoglu B., Ocak I., and Hasenekoglu İ. (2006) The accelerating effects of the microorganisms on biodeterioration of stone monuments under air pollution and continental-cold climatic conditions in Erzurum, Turkey. *Science of the Total Environment* Vol.364, 272-283.
- Pochon, J., and Jaton, C. (1968): *Biological factors in the alteration of stone*. In: A. H. Wolters, C. C. Elphick, Eds., *Biodeterioration of Materials*, Elsevier, Amsterdam, 258-268.
- Praderio G, Schiraldi A, Sorlini C, Stassi A, and Zanardini E. (1993) Microbiological and calorimetric investigation on degraded marbles from the Ca' D'Oro facade (Venice). *Thermochim Acta*, Vol.227, 205-13.
- Saiz-Jimenez C. (1997) Biodeterioration vs biodegradation: the role of microorganisms in the removal of pollutants deposited onto historic buildings. *Int Biodeterior Biodegrad*, Vol.40, 225-32.
- Samson, R.A., Houbraken J., Thrane U., Frisved J.C. and Andersen, B. (2010) *Food and Indoor Fungi* Cbs. Knaw. Fungal Biodiversity Centre Utrecht, the Netherlands an Institute of the Royal Netherlands Academy of Arts and Science.
- Smith, O. and Onions, A.H.S. (1994) *Preservation and Maintenance of Living Fungi*. CAB International, UK.
- Suihko, L. M., Alakomi, L. H., Gorbushina, A. A., Fortune, I., Marquardt, and Saarela, M. (2007) Characterization of Aerobic Bacterial and Fungal Microbiota on Surfaces of Historic Scottish Monuments. *Syst. Appl. Microbiol.*, Vol.30, 494-508.
- Tomaselli L, Lementi G, Bosco M, and Tiano P. (2000) Biodiversity of photosynthetic micro-organisms dwelling on stone monuments. *Int Biodeterior Biodegrad*, Vol.6, 251-8.
- van der Gast CJ, Knowles CJ, Wright MA, Thompson IP. (2001) Identification and characterisation of bacterial populations of an in-use metal-working fluid by phenotypic and genotypic methodology. *Int. Biodeterior. Biodegrad.*, Vol.47, 113-23.
- Warscheid Th, and Braams J. (2000) Biodeterioration of stone: a review. *Int. Biodeterior. Biodegrad.*, Vol.46, 343-63. Zanardini E, Abbruscato P, Ghedini N, Realini M, Sorlini C. (2000) Influence of atmospheric pollutants on biodeterioration of stone. *Int. Biodeterior. Biodegrad.*, Vol.45, 35-42.
- Zurita YP, Cultrone G, Sanchez-Castilo P, Sebastain E, and Boliver FC. (2005) Microalgae associated with deteriorated stonework of the fountain of Bibataun in Granada, Spain. *Int. Biod. Biodegr.* Vol.55, 55-61.